01/06/16

Marguerite and Fatma has arrived

PLASMID LOSS

renew LB media 20mL with 10µL of pre-culture.

06/06/16

PLASMID LOSS

Renew LB media. Using Marie's protocol, we prepared 2 plates for temporal quantification.

07/06/16

PLASMID LOSS

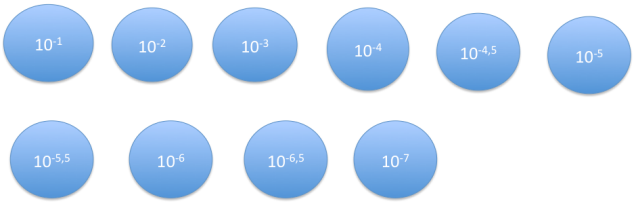
Denis advised us: Made serial dilution (in small wells) from saturated solution.
Placed onto LB media

08/06/2016

PLASMID LOSS

It seems that colonies are countable from a concentration of 10^-4. For a dilution of 10^-7, there is no colony. Dilution between 10^-4 and 10^-7 are need to see if countable colonies grows.

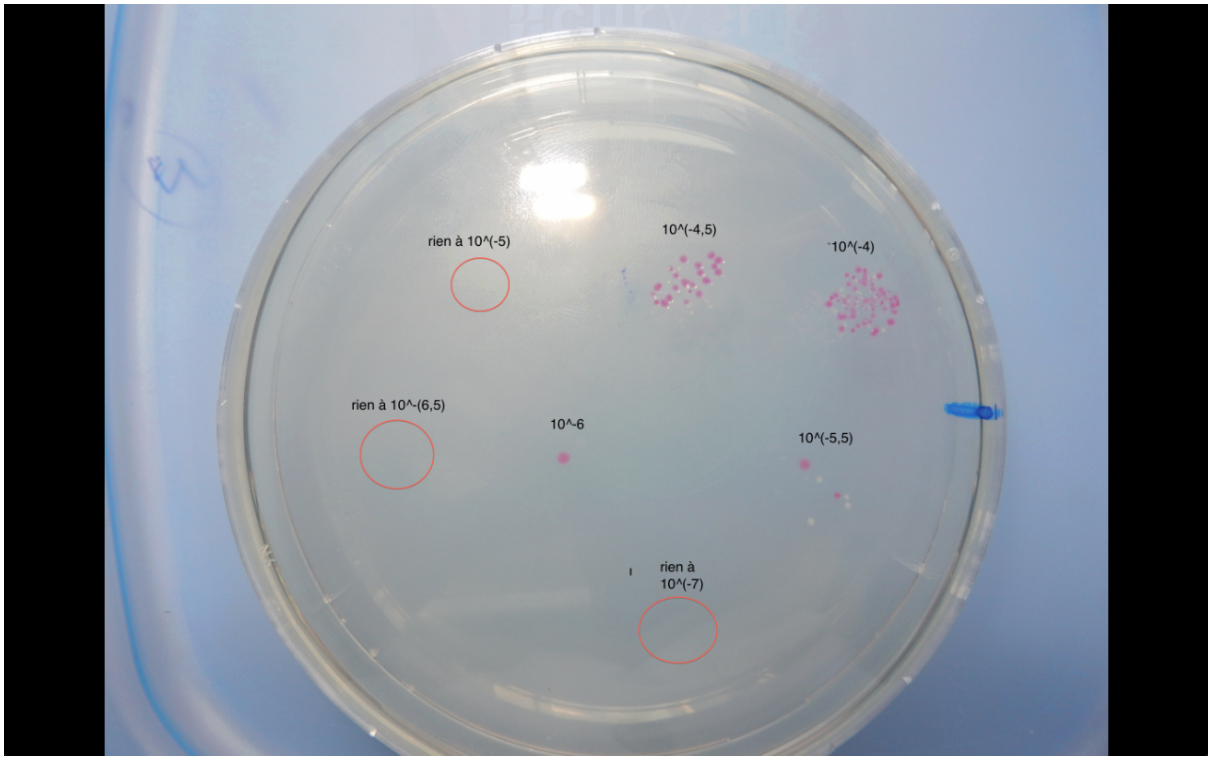
Others dilution are made. These are the folowing dilution:



09/06/2016

PLASMID LOSS

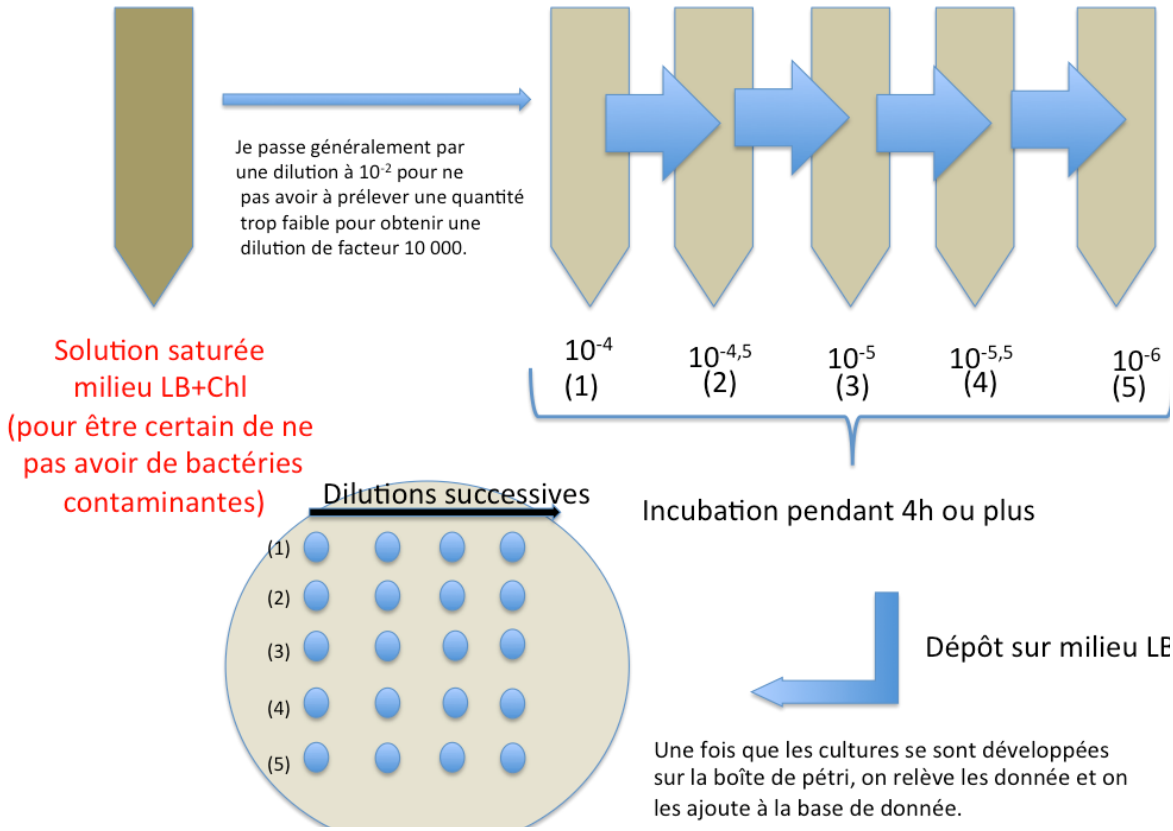
LB plate with chloramphenicol



There is some plasmid loss. But the media from which we pick out is old (few weeks). Maybe it has been contaminated.

So we picked out a pink colony and put in into a LB media + Chloramphenicol in incubation. Then, we makes sure that it isn't contaminated.

New protocol:

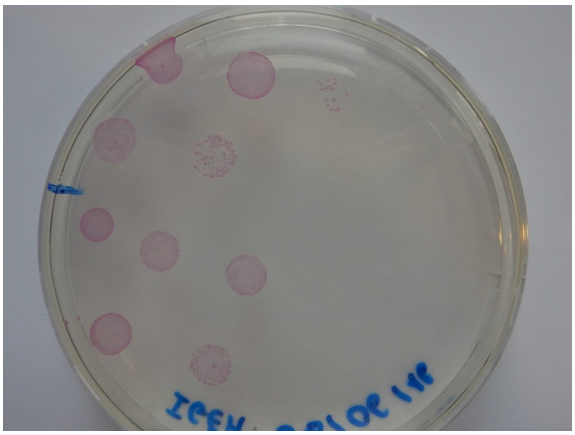


Bioinformaticians visit : Kristina, Lisa et Savandara comes to talk about the model. Bacteria have an exponential growth, we want to modelize the growth gap between bacteria with their plasmid and the ones that lose their plasmid.

Our goal : to determine a perfect bacteria concentration to have the possibility to see bacteria which lost their plasmid.

10/06/2016

PLASMID LOSS



Beginning with the new protocol with previous culture.
We will begin the culture with antibiotic tomorrow.

Plate with LB and LB+Chloramphenicol

13/06/2016

PLASMID LOSS

Previous protocol is not viable. If we want to calculate the frequency for plasmid loss, we need to know the number of cells put in culture and incubation at T=0min. But we did a serial dilution from the saturated solution, more or less saturated, we can't know the number of bacteria put in culture at T=0.
We need to make markers. We are thinking about a new protocol.

17/06/2016

PLASMID LOSS

How to make a new protocol. We set an reference OD in order to have a marker for dilution at T=0.
We wish to start for an OD of 0.1. The perfect dilution seems to be factor 1000 or 10 000.